

After treatment of the labeled glycogen with β amylase, the radioactivity was recovered in the liberated maltose.

While G-6-P increased the activity of yeast glycogen synthetase, the stimulation obtained was usually less than 2-fold (see Table II) as compared with 15- to 20-fold in the case of the animal enzymes^{1,2}.

A full report of these findings will be published elsewhere.

This investigation was supported in part by a research grant (No. G-3442) from the National Institutes of Health, U.S. Public Health Service and by the Rockefeller Foundation. The authors wish to express their gratitude to Dr. LUIS F. LELOIR for his continued interest and support of this work, and to their colleagues at the Instituto de Investigaciones Bioquímicas for valuable criticism. Special thanks are due to Dr. H. CARMINATTI for his help in the preparation of radioactive UDPG and in some preliminary experiments.

*Instituto de Investigaciones Bioquímicas Fundacion Campomar, and I. D. ALGRANATI**
Facultad de Ciencias Exactas y Naturales, E. CABIB
Obligado 2490, Buenos Aires (Argentina)

¹ L. F. LELOIR AND S. H. GOLDBERG, *J. Biol. Chem.*, 235 (1960) 919.

² L. F. LELOIR, J. M. OLAVARRIA, S. H. GOLDBERG AND H. CARMINATTI, *Arch. Biochem. Biophys.*, 81 (1959) 508.

³ J. C. TRIVELLONI, *Arch. Biochem. Biophys.*, in the press.

⁴ S. PEAT, J. R. TURVEY AND G. JONES, *J. Chem. Soc.*, (1959) 1540.

⁵ E. CABIB AND L. F. LELOIR, *J. Biol. Chem.*, 231 (1958) 259.

* Fellow of the University of Buenos Aires.

Received July 21st, 1960

Biochim. Biophys. Acta, 43 (1960) 141-142

The identification of lysolecithin in lipid extracts of brain

The presence of lysolecithin in lipid extracts of a number of mammalian tissues has recently been reported¹⁻⁵. Its presence in extracts of brain has not, as far as we are aware, been described, although PIGHINI⁶ in 1932 had obtained an alcohol-soluble, ether-insoluble substance from human brain which, both from its solubility properties and from certain of its biological actions, appeared to resemble a lysophosphatide. It was decided therefore to re-investigate PIGHINI's work, using more modern chromatographic methods for the separation of lysolecithin. Initial experiments showed that lipid extracts of normal human brain, prepared by PIGHINI's method, contained a haemolytic substance having the same R_F value as lysolecithin. We therefore next decided to extract the brain lipids by the method of FOLCH *et al.*⁷ and to characterise this haemolytic substance more precisely.

Human brains were obtained post-mortem as soon as possible after death from cases in which there had been no clinical evidence of nervous disorder. Meninges were removed and the surface of the brain washed free from blood in gently running tap-water. Cerebral cortical grey matter was then dissected off and extracted. In addition, a number of scattered plaques were dissected from the unfixed brain of a case of multiple sclerosis.

One specimen of a normal human frontal pole was obtained post-operatively, immediately placed in a vacuum flask at 0° , brought to the laboratory and treated as above. Whole rat brains were also used, and extracted immediately after killing by decapitation.

The choline-containing phosphatide fractions were separated from the total lipid extracts⁷ on Al_2O_3 columns and then applied to silicic acid-impregnated papers⁸. Chromatograms were developed with $\text{CHCl}_3\text{--CH}_3\text{OH}$ (4:1, v/v). Egg lysolecithin⁹ was used as a marker.

Selected lanes were cut from each paper, stained with phosphomolybdic acid, washed and reduced with SnCl_2 ¹⁰. Areas corresponding to the lysolecithin spots were cut out from unstained lanes and eluted with $\text{CH}_3\text{OH--CHCl}_3$ (5:1, v/v). The eluates were evaporated in a rotary evaporator at 40° and dissolved for analysis in a known volume of CHCl_3 .

Total P and P in the alkali-labile phospholipids (following hydrolysis by the method of SCHMIDT *et al.*¹¹) was determined as described previously¹². Fatty acids were determined titrimetrically after hydrolysis of lipid samples for 2 h in 2 N NaOH, followed by acidification and extraction of the fatty acids with ether. Choline was estimated by the method of WEBSTER¹³, and haemolytic activity tested by the procedure of COLLIER¹⁴.

Paper chromatograms of the choline phosphatides obtained from human brain have regularly shown the presence of 3 spots staining with phosphomolybdic acid, the slowest of which has an R_F resembling that of lysolecithin. Elution of this slowest spot from unstained papers, followed by re-chromatography on paper, showed the presence of a single spot with R_F corresponding to that of lysolecithin (Fig. 1).



Fig. 1. Chromatogram of human brain lysolecithin. A = Brain lysolecithin. B = Brain lysolecithin + added egg lysolecithin. C = Egg lysolecithin.

After elution from unstained papers this substance was in each experiment found to be haemolytic and to give total P:alkali-labile lipid P:choline:fatty acid ratios close to 1:1:1:1 (Table I). These data indicate to us therefore that total lipid extracts of brain contain appreciable amounts of lysolecithin. Confirmatory evidence of a preliminary nature has been obtained by infrared spectroscopy kindly carried out by Dr. W. L. G. GENT. It is possible of course that the lysolecithin in these extracts may have been formed in the brain post-mortem, or during the chemical manipulations involved in its isolation. It has, however, been isolated from the post-operative specimen of human brain in amounts comparable to those present in the post-mortem material, and is also present in rat brain extracted within 3 min of killing. It has also been shown that storage of the total lipid extract at -10° for 48 h causes no apparent increase in the size of the lysolecithin spot; furthermore, when lecithin was taken through the entire procedure of extraction and chromato-

TABLE I
ANALYTICAL DATA ON THE ELUATES

Tissue	$\mu\text{moles/ml eluate solution}$				Molar ratios (A:B:C:D)
	A Total P	B Alkali-labile lipid P	C Choline	D Fatty acids	
Rat brain	0.31	0.29	0.30	—	1 : 0.93 : 0.98 : —
Rat brain	4.57	4.43	4.57	4.56	1 : 0.97 : 1.00 : 1.00
Rat brain	7.06	7.48	7.01	6.65	1 : 1.06 : 0.99 : 0.94
Post-operative human brain	0.47	0.41	0.48	—	1 : 0.87 : 1.02 : —
Post-mortem human brain	7.10	6.80	6.00	6.40	1 : 0.96 : 0.85 : 0.90
Post-mortem human brain	1.44	1.44	1.51	1.52	1 : 1.00 : 1.05 : 1.06
Post-mortem human brain	1.60	1.52	1.62	1.70	1 : 0.95 : 1.01 : 1.06
Post-mortem human brain	0.86	0.90	0.88	0.93	1 : 1.05 : 1.02 : 1.08
Post-mortem human brain	1.51	1.54	1.57	1.61	1 : 1.02 : 1.04 : 1.07
Plaque tissue from brain of case of multiple sclerosis	0.89	0.92	0.90	—	1 : 1.03 : 1.01 : —

graphic separation used in the brain experiments no evidence of lysolecithin formation was observed.

Although our methods are not yet entirely suitable for quantitative work (and for this reason the data in Table I are expressed as $\mu\text{mole/ml eluate solution}$), the amounts of lysolecithin present would appear to correspond to 1–4 $\mu\text{moles/g}$ fresh tissue. Further work is now in progress to put these observations on a quantitative basis.

Our thanks are due to Mr. WYLIE MCKISSOCK, F.R.C.S., for providing us with the post-operative sample of human brain and to Miss JUDITH FIDLER for her technical assistance.

Department of Chemical Pathology, Guy's Hospital Medical School, R. H. S. THOMPSON
London, S.E. 1 (Great Britain) R. NIEMIRO*
G. R. WEBSTER

¹ G. V. MARINETTI, R. F. WITTER AND E. STOTZ, *J. Biol. Chem.*, 226 (1957) 475.

² S. HAJDU, H. WEISS AND E. TITUS, *J. Pharmacol. Exptl. Thermol.*, 120 (1957) 99.

³ G. B. PHILLIPS, *Proc. Natl. Acad. Sci., U.S.A.*, 43 (1957) 566.

⁴ E. GJONE, J. F. BERRY AND D. A. TURNER, *J. Lipid Research*, 1 (1959) 66.

⁵ H. P. SCHWARZ, L. DREIBACH, R. STAMBAUGH, A. KLESCHICK AND M. BARRIONUEVO, *Arch. Biochem. Biophys.*, 87 (1960) 171.

⁶ G. PIGHINI, *Z. ges. Neurol. Psychiat.*, 140 (1932) 218.

⁷ J. FOLCH, M. LEES AND G. H. SLOANE-STANLEY, *J. Biol. Chem.*, 226 (1957) 497.

⁸ C. H. LEA, D. N. RHODES AND R. D. STOLL, *Biochem. J.*, 60 (1955) 353.

⁹ E. A. MARPLES, R. H. S. THOMPSON AND G. R. WEBSTER, *J. Neurochem.*, 4 (1959) 62.

¹⁰ E. CHARGAFF, C. LEVINE AND C. GREEN, *J. Biol. Chem.*, 175 (1948) 67.

¹¹ G. SCHMIDT, J. BENOTTI, B. HERSHMAN AND S. J. THANNHAUSER, *J. Biol. Chem.*, 166 (1946) 505.

¹² K. P. STRICKLAND, R. H. S. THOMPSON AND G. R. WEBSTER, *J. Neurol. Neurosurg. Psychiat.*, 19 (1956) 12.

¹³ G. R. WEBSTER, *Biochim. Biophys. Acta*, 20 (1956) 432.

¹⁴ H. B. COLLIER, *J. Gen. Physiol.*, 35 (1952) 617.

* Polish Government Overseas Research Fellow.

Received July 8th, 1960